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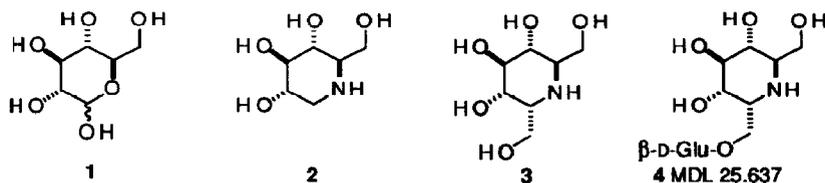
Synthesis of Aza-C-disaccharides - a New Class of Sugar Mimics

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Abstract: Synthesis of a novel aza-C-disaccharide is described using a Suzuki coupling of the alkyl boron reagent **8** and vinyl bromide **7** as the key step.

Azasugars have attracted much interest due to their diverse structure and biological activity.¹ They have potential in the treatment of diabetes and other metabolic disorders,² cancer³ and some viral diseases (notably human immunodeficiency virus (HIV)).⁴

The most widely recognized azasugars mirror the structure of the most common hexoses found in nature, *cf.* D-glucose (**1**) and 1-deoxynojirimycin (**2**), and are strong inhibitors of *exo*-glycosidases.^{1, 5} The hepta-azasugar homologs that contain a hydroxymethyl substituent at the pseudo-anomeric carbon (*e.g.*, α -1-homonojirimycin (**3**)) display stronger selectivity for inhibition of α - vs. β -*exo*-glycosidases and *endo*-glycosidases.^{2b,2c,6} Presumably the hydroxymethyl group mimics the second sugar unit of a disaccharide which results in a more efficient glycosidase inhibitor. A potent and selective glycosidase inhibitor and an antidiabetic drug candidate has been produced by carrying this analogy one step further with the synthesis of MDL 25,637 (**4**) in which a second sugar is linked by a β -glycosidic bond.^{2b,c} Such β -glycosides are generally more stable towards hydrolysis by pertinent mammalian enzymes.

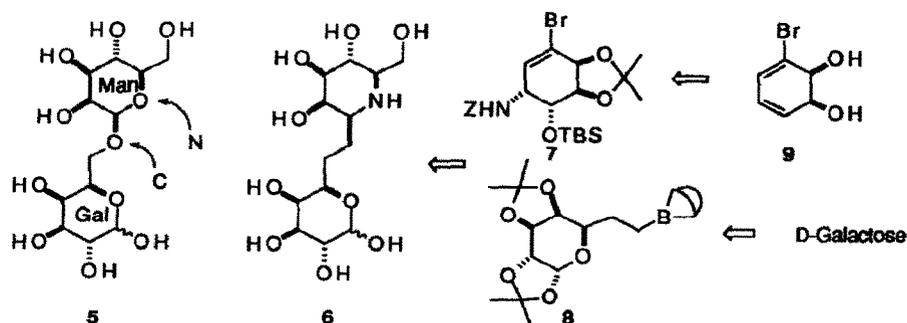
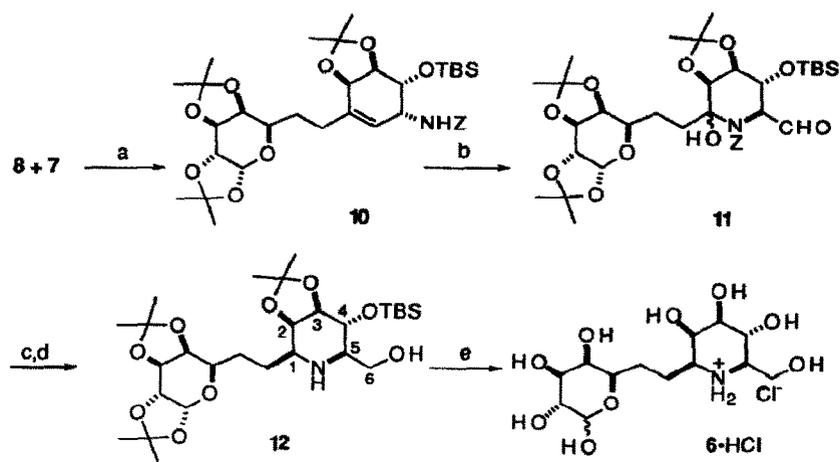


The synthesis of oligosaccharide analogues containing various glycoside-linked 1-deoxynojirimycins and related moieties at the reducing end have been reported.⁷ The preparation of an azapyranosyl thioglycoside having the azasugar residue at the non-reducing end has also been described.⁸

In this communication, we present a model synthesis of the aza-C-disaccharide **6** (Scheme I). This novel sugar mimic, fully resembling a parent disaccharide, 6-O- β -D-mannopyranosyl-D-galactose (**5**), should be inert towards acidic or enzymatic hydrolysis while preserving strong binding properties of the parent azasugar, 1-deoxy-*manno*-nojirimycin. Replacement of the glycosidic oxygen by a methylene unit should not significantly affect the conformation of the glycosidic linkage as illustrated by Kishi and coworkers.⁹

Vinyl bromide **7** was prepared according to Hudlicky's procedure¹⁰ from enantiopure bromo diol **9**,¹¹ a product from microbial oxidation of bromobenzene. The alkyl boron reagent **8** was generated by hydroboration (1.5 eq of 9-BBN, THF, 4 h) of the corresponding olefin¹² derived in four steps from D-galactose. The key transformation was achieved using Suzuki coupling¹³ of **7** and **8** in 80% yield. Ozonolysis followed by reductive work-up gave the keto-aldehyde, which existed predominately in the cyclic form **11** based on its ¹H NMR spectrum (Scheme II).

Scheme I

Scheme II^a

^aReagents and conditions: a) PdCl₂(dppf), DMF, K₃PO₄, rt (80%); b) O₃, MeOH:CH₂Cl₂ (1 : 1), -78 °C, then DMS 1 h; c) NaBH₃CN, MeOH, pH-4 buffer, rt; d) H₂, Pd-C, MeOH (57 % from 10); e) 1 N HCl, rt (95 %).

Chemoselective reduction of the aldehyde of 11 was achieved using sodium cyanoborohydride-HOAc/NaOAc buffer (borane-*t*-butyl amine complex in dichloromethane gave comparable results) (Scheme II). Hydrogenation of the resulting amination gave the β-pseudo anomer 12 as a single diastereomer based on ¹H NMR (500 MHz) analysis of the crude product (57% isolated yield from the alkene 10).¹⁴ The relative stereochemistry of the pseudo-anomeric center was established on the coupling pattern as well strong NOE effect between H-1 and H-5 protons (Figure 1). Acidic deprotection of all blocking groups afforded the hydrochloride salt of 6 as an anomeric mixture in 95 % yield (α : β; 1 : 2).

An efficient method for the production of 1-6 linked aza-C-disaccharides has been developed. The scope and limitations of this methodology and the relevant biological activities of this new class of compounds will be reported in due course.

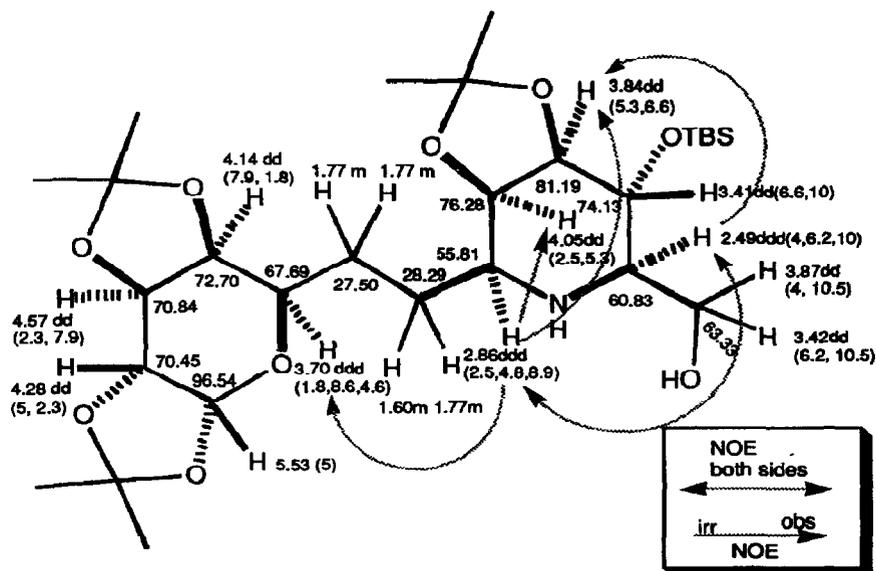


Figure 1. ^1H NMR data for compound 12: Proton assignments by ^1H decoupling and DDR. Carbon assignments by HETCOR experiments. Relative stereochemistries by NOE experiments. All data obtained in CDCl_3 at 500 MHz.

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14. Selected Physical Data:
10: $[\alpha]_D$ -57.5 (c 0.8, CH₂Cl₂); ¹H NMR (CDCl₃, 300 MHz) δ 0.05 (s, 3 H), 0.08 (s, 3 H), 0.85 (s, 9 H), 1.31 (s, 3 H), 1.34 (s, 6 H), 1.36 (s, 3 H), 1.46 (s, 3 H), 1.51 (s, 3 H), 1.67 (m, 1 H), 1.88 (m, 1 H), 2.16 (m, 1 H), 2.34 (m, 1 H), 3.71 (ddd, 1 H, J = 1.5, 4.2, 5.7 Hz), 4.09 (dd, 1 H, J = 1.8, 8.1 Hz), 4.18 (m, 2 H), 4.28 (dd, 1 H, J = 2.4, 5.4 Hz), 4.42 (d, 1 H, J = 4.8 Hz), 4.47 (d, 1 H, J = 8.7 Hz), 4.56 (dd, 1 H, J = 2.4, 8.1 Hz), 4.94 (d, 1 H, J = 9.6 Hz), 5.12 (s, 2 H), 5.30 (d, 1 H, J = 1.8 Hz), 5.53 (d, 1 H, J = 5.1 Hz), 7.40 (m, 5H); ¹³C NMR (CDCl₃, 75.5 MHz) δ -5.01, -4.78, 17.87, 24.32, 24.80, 25.66, 25.99, 26.16, 26.55, 27.72, 27.82, 28.83, 48.17, 66.56, 67.03, 69.93, 70.46, 70.91, 72.87, 73.44, 75.81, 96.46, 108.10, 108.88, 109.38, 122.11, 127.95, 128.38, 136.62, 137.70, 155.61.
12: $[\alpha]_D$ -36.1 (c 2.5, CH₂Cl₂).

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